

Structure and Stereochemistry of 4 α ,7 α ,7 β -Nepetalactone from *Nepeta mussini* and Its Relationship to the 4 α ,7 α ,7 α - and 4 α ,7 α ,7 β -Nepetalactones from *N. cataria*

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The structure of the major constituent of the steam-volatile oil from *Nepeta mussini* has been established, through NMR spectroscopy and chemical correlation, to be that of a methylcyclopentane monoterpene diastereomeric with the two nepetalactones from *N. cataria*, all having the same absolute configuration at C-7. Stereochemical assignments to the four nepetalactones derived from these nepetalactones are also reported.

4 α ,7 α ,7 α -Nepetalactone (**1a**), isolated from *Nepeta cataria*,² was the first methylcyclopentane monoterpene to be fully characterized.³ Subsequently, numerous such monoterpenoids of plant and animal origin were identified, many by correlation with **1a** and its degradation products.⁴ In addition to **1a**, *N. cataria* produces, as a minor component, a second diastereomeric enol lactone, 4 α ,7 α ,7 β -nepetalactone (**1b**),^{5a} which has been purified^{5b} to the crystalline state, mp 37–39 °C, in the current work by high-pressure liquid chromatography (high-pressure LC). This paper describes the isolation, structure proof, physical properties, and spectral data of a third diastereomeric enol lactone, 4 α ,7 α ,7 β -nepetalactone (**1c**), from *N. mussini*.⁶ A comparison of these three nepetalactones and their respective degradation products is made.

The major constituent of the steam-volatile oil from the foliage of *N. mussini* was previously reported to be lactone **1b**.⁷ However, subsequent chemical degradation of this lactone^{1c,d} showed that this assignment was incorrect and should be that of **1c**. The spectra of **1a–c** and their degradation products were examined to confirm the nonequivalence of these enol lactones and to establish the structure of **1c** from *N. mussini*.

The steam-volatile oil obtained from *N. mussini* contains approximately 95% **1c** and only minor amounts of **1a** as indicated by high-pressure LC and ¹³C NMR. Chloroform extracts of macerated fresh *N. mussini* foliage that had not been steam distilled or even heated showed the same ratio of **1c** to **1a** when examined by high-pressure LC. Preparative high-pressure LC of the steam-volatile oil gave pure **1c**. Lactones **1a** and **1b** were similarly isolated by high-pressure LC from the steam-volatile oil of *N. cataria*.

Table I. ¹³C NMR Chemical Shifts for Nepetalactones **1a–c**^{a,b}

carbon	1a	1b	1c
1	170.5	169.9	169.8
3	133.4	135.7	134.0
4	115.1	120.4	115.4
4a	(40.7)	(37.3)	(39.4)
5	(30.9)	(26.1)	(30.4)
6	(33.0)	(30.0)	(32.7)
7	(39.7)	(32.1)	(38.3)
7a	49.3	49.0	46.2
8	15.4	14.3	14.7
9	20.3	17.6	17.2

^a Spectra were recorded as 0.6 M solutions in CDCl₃; chemical shifts are reported in parts per million downfield from internal Me₄Si. ^b Parentheses indicate uncertain assignments, as between C-4a and C-7 and as between C-5 and C-6.

Table II. Selected ¹H NMR Chemical Shifts for Nepetalactones **1a–c**^a

proton	1a	1b	1c
3 (vinyl)	6.15 (nm) ^b	6.23 (nm)	6.18 (nm)
8 (allylic CH ₃)	1.64 (nm)	1.71 (nm)	1.70 (nm)
9 (CH ₃)	1.19 (d, 6) ^b	1.11 (d, 6)	0.99 (d, 7)

^a Spectral conditions same as in Table I; values in parts per million downfield from internal Me₄Si. ^b Abbreviations in parentheses indicate narrow multiplet (nm) or doublet (d) followed by coupling constant in hertz.

The assignment of configuration of **1c** is based on its ¹³C and ¹H NMR spectra. Chemical shifts (¹³C and ¹H) for **1a**, **1b**, and **1c** are listed in Tables I and II. Assignments of peaks in the ¹³C spectra are from off-resonance decoupled ¹³C spectra of **1a** and **1c** (peaks for **1b** assigned by analogy), distinguishing C-5/C-6 from C-4a/C-7 and C-7a as well as C-8/C-9 and based on known chemical-shift effects (C-1, -3, and -4). The C-7a signal is downfield from peaks for the other methine carbons, owing to the adjacent carbonyl group; C-4a and C-7 cannot readily be distinguished, nor can C-5 and C-6. The assignment of signals for C-8 and C-9 is discussed below.

The ¹³C spectrum of **1c** (Table I) closely resembles that of **1a** (cis ring junction) but not that of **1b** (trans ring junction). Thus, **1c** must be the other cis diastereomer. The only sizeable differences between **1a** and **1c** appear at C-9 and C-7a.⁸ The carbonyl (C-1 group) is cis to C-9 in **1c** and trans in **1a**, and accordingly its signal is upfield

(8) One might have expected differences at C-1 and C-7 also. Shifts due to cis–trans isomerism of carbonyl substituents often tend to be attenuated.⁹ The shift at C-7 is present but small.

(9) E. L. Eliel and K. M. Pietrusiewicz, *Org. Magn. Reson.*, **13**, 193 (1980).

(1) (a) I am grateful to the chemistry faculty at Chapel Hill for their hospitality during a 1977 sabbatical visit. (b) Oklahoma State University, Stillwater. (c) Ph.D. Dissertation, Oklahoma State University, Stillwater, OK, 1974. (d) Ph.D. Dissertation, Oklahoma State University, Stillwater, OK, 1968. (e) University of North Carolina, Chapel Hill.

(2) (a) S. M. McElvain, P. M. Walters, and R. D. Bright, *J. Am. Chem. Soc.*, **64**, 1828 (1942); (b) J. Meinwald, *ibid.*, **76**, 4571 (1954).

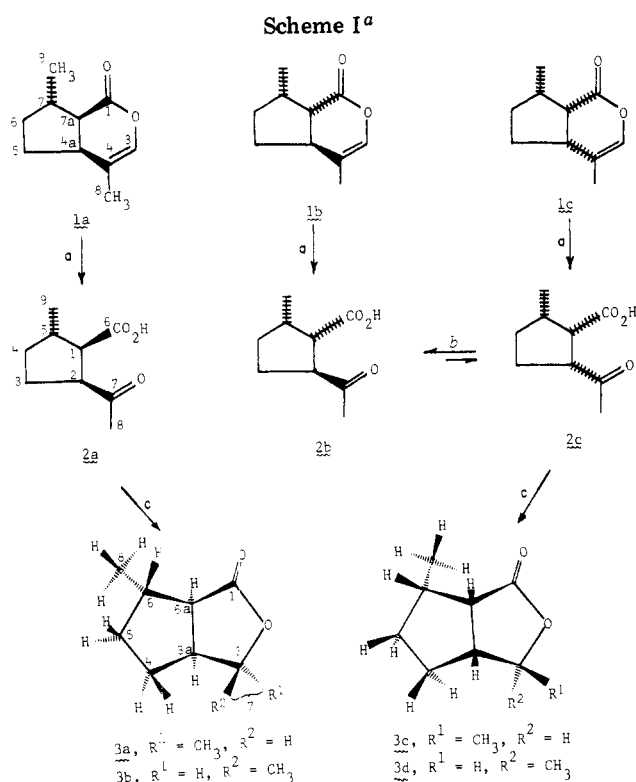
(3) (a) S. M. McElvain and E. J. Eisenbraun, *J. Am. Chem. Soc.*, **77**, 1599 (1955); (b) E. J. Eisenbraun and S. M. McElvain, *ibid.*, **77**, 3383 (1955); (c) S. M. McElvain and E. J. Eisenbraun, *J. Org. Chem.*, **22**, 976 (1957); (d) R. B. Bates, E. J. Eisenbraun, and S. M. McElvain, *J. Am. Chem. Soc.*, **80**, 3420 (1958).

(4) W. I. Taylor and A. R. Battersby, Eds., "Cyclopentanoid Terpene Derivatives", Marcel Dekker, Inc., New York, 1969.

(5) (a) R. B. Bates and C. W. Sigel, *Experientia*, **19**, 564 (1963). (b) Since the specific rotation of **1b** from the earlier purification^{5a} was reported as +20.2° and calculated as +21.9° for a pure sample, in contrast to [α]_D²⁵ –24.4° (c 6.15, CHCl₃) for our crystalline sample, we conclude that a typographical error resulted or that the current preparation is the first description of pure **1b**.

(6) R. Trave, A. Marchesini, and L. Garanti, *Gazz. Chim. Ital.*, **98**, 1132 (1968).

(7) F. E. Regnier, G. R. Waller, and E. J. Eisenbraun, *Phytochemistry*, **6**, 1281 (1967).



^a a, O₃, CH₃OH, pyridine; b, OH⁻, H₃O⁺; c, NaBH₄.

in **1c** and downfield in **1a** (the methyl shift in *cis*-1,2-dimethylcyclopentane is 16.1, that in the *trans* isomer 19.7 ppm).¹⁰ The assignments of C-8 (nearly constant for the three diastereomers) and C-9 were made accordingly. Substitution at C-7a is *cis-cis* to that at C-7 and C-4a in **1c** but *trans-cis* in **1a** and **1b**; since the signals of the substituted ring carbons in *cis*-1,2-dimethylcyclopentane (38.6 ppm) are upfield from those in the *trans* isomer (43.7 ppm), it is reasonable that the C-7a signal in **1c** (46.2 ppm) is upfield from that for **1a** (49.3 ppm) as well as that for **1b** (49.0 ppm).

The ¹³C spectrum of **1b** differs notably from the other two at C-4, C-4a, C-5, C-6, and C-7, whereas, for reasons already mentioned, it resembles **1a** at C-7a, **1c** at C-9, and both diastereomers at C-8. The downfield shift at C-4 stems from its being *trans* to C-1—*vide supra*. The upfield shifts at the other four positions (4a, 5, 6, 7) may be explained as follows: the *trans* junction of the five-membered ring to the six-membered ring leads to appreciable puckering of the former. The C-9 methyl group in **1b** thus occupies an "axial-like" position unlike those in **1a** and **1c**. This will bring into effect upfield-shifting γ_a effects (at C-5 and C-4a) and differentially upfield-shifting¹¹ α_a (at C-7) and β_a effects (at C-6; it is not clear why this differential β effect is not also seen at C-7a).

The *cis* arrangement of the protons of the ring junction of **1c** is also evident from the ¹H NMR spectrum (Table II). The C-9 methyl proton signals in **1c** are shifted markedly upfield relative to those in **1a** because the methyl group in **1c** extends over the face (shielding region) of the carbonyl group; to a much lesser degree this is also true for **1b**.

In view of the suspected stereochemical relationship of **1c** to **1a** or **1b**, the obvious chemical proof should come

(10) M. Christl, H. J. Reich, and J. D. Roberts, *J. Am. Chem. Soc.*, **93**, 3463 (1971).

(11) In the sense that the α_a and β_a effects are less downfield shifting than α_e and β_e.

Table III. ¹³C NMR Chemical Shifts for Nepetalactones **3a-d**^a

carbon	3a	3b	3c	3d
1	179.8	179.2	177.6	177.2
3	82.4	76.8	82.4	77.1
3a	47.0	44.5	47.9	43.5
4	31.4	25.7	31.6	25.2
5	34.3	35.1	33.9	34.4
6	39.0	38.3	38.1	38.2
6a	52.6	54.2	48.6	49.9
7	22.2	16.4	22.2	16.6
8	20.5	21.2	15.8	15.3

^a Conditions same as in Table I; shifts are in parts per million downfield from internal Me₄Si.

from correlation of structure through degradation to a common intermediate of established structure and configuration. We proceeded as shown in Scheme I with degradation of **1c** through ozonolysis to nepetonic acid (**2c**). Nepetonic acid (**2c**) displays tautomeric behavior (lactol formation) as evidenced by line broadening in the ¹H NMR spectrum and hydroxyl group absorption^{3d} in the IR spectrum. Treatment of **2c** with diazomethane yields a single oxo ester which readily forms a yellow 2,4-dinitrophenylhydrazone (2,4-DNP), mp 122–123 °C.

As expected, nepetonic acid (**2c**) is unstable relative to *trans*-nepetonic acid (**2b**), to which it is readily epimerized. Treatment of the epimerization product with CH₂N₂ gave two methyl esters which were shown by GC analysis to be **2b** and **2c** in a 94:6 ratio. This ester mixture was identical with one similarly prepared from **1b**, as shown by their ¹H NMR and IR spectra and the specific rotation of the parent esters as well as their 2,4-DNP derivatives, mp 75–77 °C and 76–78 °C, undepressed by admixture. This correlation clearly relates **1b** and **1c** as being diastereomers epimeric at C-4a but having identical absolute configuration at C-7 and C-7a. The absolute configuration of C-7 of **1b** is known through its relationship to **1a**, which in turn had been degraded^{3a} to (–)-3-methylcyclopentanone and (–)-methylsuccinic acid of known absolute configuration.^{3b} Further, **1a** has been chemically related to isoiridomyrmecin,^{3c} for which the structure was established through X-ray crystallography.¹²

The nepetalactones **3a** and **3b** have been known ever since the structure of nepetalactone **1a** was established.^{3a} they were originally obtained as coproducts in the preparation of nepetonic acid **2a** from nepetalactone **1a** through alkaline hydrogen peroxide oxidation. Their skeletal system demands a *cis* ring junction;¹³ consequently the isolation of the new pair of nepetalactones **3c** and **3d** as degradation products of **1c** points to a *cis* ring junction for **1c** also. This argument is compelling when it is considered that these new nepetalactones are derived from alkali-labile nepetonic acid **2c**, as shown in Scheme I.

The ¹³C NMR spectra of **3a-d** (Table III)¹⁴ are in ex-

(12) J. F. McConnell, A. McL. Mathieson, and B. P. Schoenborn, *Tetrahedron Lett.*, 445 (1962).

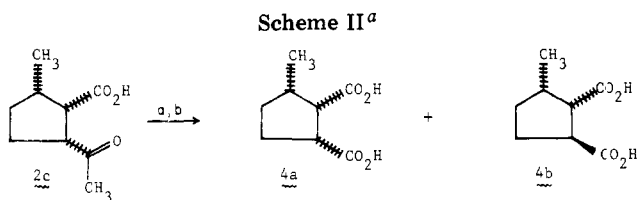
(13) The γ-hydroxy acids obtained through NaBH₄ reduction of **2a** or **2c** spontaneously lactonize at pH 7–8, whereas the γ-hydroxy acids obtained from **2b** or other *trans*-nepetonic acids do not lactonize on treatment with acid or distillation.^{3a}

(14) Off-resonance decoupling distinguished carbons 3/3a/6/6a, 4/5, and 7/8. Shifts for C-1 and C-3 are assigned by chemical-shift analogy. The signal for C-6a is downfield from those of C-6 and C-3a because of the adjacent carbonyl group, and the C-3a peak is downfield from that for C-6 because of the larger number of β substituents. The C-5 signal is identified from its expected and observed constancy among the four diastereomers. The C-7 and C-8 signals are assigned to maintain constancy of the C-8 shift between **3a** and **3b** (as well as **3c** and **3d**) and of the C-7 shift between **3a** and **3c** (or **3b** and **3d**).

Table IV. Selected ^1H NMR Chemical Shifts for Nepetalactones 3a-d^a

proton	3a	3b	3c	3d
3	4.31 (m) ^b	4.60 (m)	4.15 (m)	4.69 (m)
7	1.36 (<i>J</i> = 6) ^b	1.30 (<i>J</i> = 6)	1.37 (<i>J</i> = 6)	1.37 (<i>J</i> = 6)
8	1.11 (<i>J</i> = 6)	1.07 (<i>J</i> = 6)	1.17 (<i>J</i> = 7)	1.24 (<i>J</i> = 7)

^a Spectral conditions same in Table I; values in parts per million downfield from internal Me₄Si. ^b (m) refers to multiplet; coupling constants are in hertz.



^a a, Br₂, OH⁻; b, H₃O⁺.

cellent agreement with the assigned structures based on the fact¹⁰ (vide supra) that methyl and α carbon signals in *cis*-1,2-dimethylcyclopentanes are upfield of corresponding signals in the *trans* isomers. As one proceeds from 3a to 3b, the C-7/C-4 relationship changes from *trans* to *cis*. This should give upfield shifts for C-7, C-3, C-3a, and C-4 in 3b relative to 3a, and indeed these shifts are observed, whereas the remaining carbon signals show very little shift. An entirely analogous situation (and for analogous reasons) is seen in the comparison of 3c (C-7/C-4 *trans*) and 3d (C-7/C-4 *cis*). On the other hand, 3a and 3c are "enantiomer-like" in the lactone region of the molecule and differ only in the C-8/C-1 relationship (*trans* in 3a, *cis* in 3c). Hence, this pair should be spectrally similar except at C-1, C-8, and the juncture points C-6 and C-6a; signals for three of these carbons are notably upfield in 3c (the effect at C-6 is very small), whereas the remaining carbons show very similar shifts for 3a and 3c. Indeed, the shifts for C-3 and C-7 are exactly the same in these two diastereomers! An exactly analogous situation applies to 3b compared to 3d.

The proton spectra (Table IV) are in accord with the assigned configurations. Isomers 3a and 3c differ markedly at H-3a from 3b and 3d, with the shift of this proton being relatively upfield when it is *cis* to Me-7 (3a, 3c) and downfield when it is *trans* (3b, 3d). Similar sizable shift differences have been reported¹⁰ between the CHOAc protons in *cis*- and *trans*-2-methylcyclopentyl acetates. Also, the protons on the Me-8 group are at higher field when the methyl is *trans* to the adjoining C=O (3a, 3b) and at lower field when it is *cis* (3c, 3d). This is probably a consequence of the well-known maximum in deshielding by the C=O moiety which occurs in the plane of the carbonyl function.

Further information about the structure, stereochemistry, and configuration of nepetic acid (2c) was gained through hypohalite degradation to the known nepetic acids 4a and 4b, as shown in Scheme II. In the haloform reaction, some 2c is epimerized at C-2 to 2b at a rate competitive with halogenation α to the oxo group. Methyl esters of these acids 4a and 4b were compared with the corresponding synthesized methyl nepetates¹⁵ through use

of GC, optical rotation, and melting point of appropriately constituted mixtures.^{1c}

Experimental Section

Proton and ^{13}C NMR spectra (fully decoupled and single-frequency off-resonance proton-decoupled) were recorded with a Varian XL-100-15 interfaced with a 32K Nicolet 1080 computer at 100.1 and 25.2 MHz, respectively; some ^{13}C spectra were recorded with a Varian XL-100-12 instrument at 25.16 MHz. Chemical shifts are reported in parts per million downfield from internal Me₄Si, with coupling constants in hertz. Pulsed FT carbon spectra were obtained from approximately 1.5 M solutions in CDCl₃ containing 5% Me₄Si (δ CDCl₃ = 76.9 ppm), using a 6000-Hz sweepwidth and 8K data points. The majority of the NMR studies were carried out at Chapel Hill.^{1a} However, we thank the National Science Foundation for Grants GP 17641 and CHE76-5571 which enabled purchase of NMR instrumentation at Stillwater. IR spectra were recorded with a Beckman IR-8 spectrometer. Mass spectra were recorded with a CEC-21-110B spectrometer at 70 eV. Optical rotations were measured with a Rudolph Model 80 polarimeter. Melting and boiling points are uncorrected. Gas chromatographic analyses were conducted with a Hewlett-Packard 5750 instrument equipped with dual flame-ionization detectors and an 8 ft \times 0.25 in. copper tube packed with 6% UC W-98 coated on 80-100-mesh AW DMCS-treated Chromosorb G. Analytical and preparative high-pressure LC were conducted on a Waters Associates Micro-Porasil (silica gel) column with a Waters Associates analytical or Prep-500 LC system equipped with UV and index of refraction detectors.

4 α ,7 α ,7 β -Nepetalactone (1c). *N. mussini* plants (674 g) grown from seed were harvested, immediately shredded in a 1-gal. Waring Blendor with 1 L of water, and subjected to steam distillation. The steam distillate was saturated with NaCl and extracted with CH₂Cl₂ (3x). The CH₂Cl₂ extracts were combined, dried (Na₂SO₄), and evaporated under vacuum. Distillation of the residue [40-80 $^{\circ}\text{C}$ (0.3 mm)] gave 4.0 g (0.6% yield) of oil. High-pressure LC analysis using CHCl₃ extracts of fresh plants indicated an identical ratio of 1a to 1c. Preparative high-pressure LC of the steam-volatile oil followed by bulb-to-bulb (Kugelrohr) distillation gave pure 1c: bp 60 $^{\circ}\text{C}$ (0.1 mm); $[\alpha]_D^{25} +81.0^{\circ}$ (c 0.038, CHCl₃); ^1H NMR (CDCl₃) δ 6.18 (narrow m, 1 H, H-2), 3.20-1.00 (7 H, CH and CH₂), 1.70 (narrow m, 3 H, H-8), 0.99 (d, *J* = 7 Hz, 3 H, H-9); IR (neat) 1765, 1140 cm⁻¹; mass spectrum, *m/e* (relative intensity) 166 (M⁺, 55), 123 (98), 95 (60), 81 (100), 68 (64).

Anal. Calcd for C₁₀H₁₄O₂: C, 72.66; H, 8.49. Found: C, 72.32; H, 8.43.

4 α ,7 α ,7 β -Nepetalactone (1a) and 4 α ,7 α ,7 β -Nepetalactone (1b). Preparative high-pressure LC of the steam-volatile oil obtained from *N. cataria* (same conditions as described for *N. mussini* oil) gave 1a and 1b. Lactone 1a: $[\alpha]_D^{21} +3.7^{\circ}$ (c 27, CHCl₃); ^1H NMR (CDCl₃) δ 6.15 (narrow m, 1 H, H-2), 2.90-1.20 (7 H, CH and CH₂), 1.64 (narrow m, 3 H, H-8), 1.19 (d, *J* = 6 Hz, 3 H, H-9); IR (neat) 1765, 1140 cm⁻¹; mass spectrum, *m/e* (relative intensity) 166 (M⁺, 33), 123 (88), 109 (38), 95 (100), 81 (39), 69 (51). Lactone 1b: mp 37-39 $^{\circ}\text{C}$; $[\alpha]_D^{21} -24.4^{\circ}$ (c 6.15, CHCl₃); ^1H NMR (CDCl₃) δ 6.23 (narrow m, 1 H, H-2), 2.80-1.20 (7 H, CH and CH₂), 1.71 (narrow m, 3 H, H-8), 1.11 (d, *J* = 6 Hz, 3 H, H-9); IR (neat) 1765, 1140 cm⁻¹; mass spectrum, *m/e* (relative intensity) 166 (M⁺, 66), 123 (79), 109 (52), 94 (70), 81 (100), 69 (100).

1 α ,2 β ,5 α -Nepetic Acid (2b) and Methyl Ester. A solution of 300 mg (1.81 mmol) of 1b in 40 mL of methanol containing 1 mL of pyridine was cooled to -70 $^{\circ}\text{C}$. Ozone was bubbled through the solution until a blue color persisted. Excess O₃ was flushed from the system with N₂ and then dimethyl sulfide (5 mL) was added. The mixture was allowed to warm to room temperature (1 h), the solvent was removed under reduced pressure, and the residue was dissolved in NaHCO₃ solution and then extracted with ether. The aqueous solution was acidified with 10% HCl and extracted with ether. The ether extract was dried (Na₂SO₄) and evaporated under vacuum to give 200 mg (98% yield) of 2b; ^1H NMR (CDCl₃) δ 9.1 (br hump, 1 H, acidic H), 3.60-3.10 (overlapping m, 2 H, H-1 and H-2), 2.60-1.00 (5 H, H-3, H-4, and H-5), 2.20 (s, 3 H, H-8 methyl), 0.98 (d, *J* = 7 Hz, 3 H, H-9 methyl).

(15) K. S. Schorno, G. H. Adolphsen, and E. J. Eisenbraun, *J. Org. Chem.*, **34**, 2801 (1969); (b) E. J. Eisenbraun, G. H. Adolphsen, K. S. Schorno, and R. N. Morris, *J. Org. Chem.*, **36**, 414 (1971).

Treatment of **2b** with excess CH_2N_2 in ether and Kugelrohr distillation gave 295 mg (89% yield based on **1b**) of the methyl ester, bp 40–60 °C (0.4 mm). Gas chromatographic analysis indicated the product is a mixture of esters, arising 99% from **2b** and 1% from **2c**: $[\alpha]_D^{25} +38^\circ$ (c 3.4, CHCl_3); $^1\text{H NMR}$ (CDCl_3) δ 3.68 (s, 3 H, CO_2CH_3), 3.50–3.10 (overlapping m, 2 H, H-1 and H-2), 2.18 (s, 3 H, H-8 methyl), 0.88 (d, $J = 6$ Hz, 3 H, H-9 methyl); IR (neat) 1745, 1725 ($\text{C}=\text{O}$) cm^{-1} ; mass spectrum, m/e (relative intensity) 184 (M^+ , 3), 141 (12), 124 (20), 109 (28), 82 (14), 81 (100). The 2,4-DNP derivative of the methyl ester of **2b** was prepared: mp 75–77 °C; $[\alpha]_D +160^\circ$ (c 5.1, CHCl_3).

Anal. Calcd for $\text{C}_{16}\text{H}_{20}\text{N}_4\text{O}_6$: C, 52.74; H, 5.53; N, 15.38. Found: C, 52.46; H, 5.50; N, 15.16.

1 α ,2 α ,5 α -Nepetonic Acid (2c) and Methyl Ester. Treatment of 420 mg (2.62 mmol) of **1c** with ozone and workup in the manner described above gave 360 mg (91% yield) of **2c**; $^1\text{H NMR}$ (CDCl_3) lactol forms present, δ 7.50 (br hump, 1 H, acidic H), 3.20–2.80 (overlapping m, 2 H, H-1 and H-2), 2.40–1.00 (5 H, H-3, H-4, and H-5), 2.00 (br humps, 3 H, H-8 methyl), 1.09 (d, $J = 7$ Hz, 3 H, H-9 methyl).

A portion of this **2c** was treated with CH_2N_2 to give the methyl ester. Gas chromatographic analysis indicated 94% **2c** and 6% **2b**: $^1\text{H NMR}$ (CDCl_3) δ 3.65 (s, 3 H, CO_2CH_3), 3.20–2.85 (overlapping m, 2 H, H-1 and H-2), 2.80–1.25 (5 H, H-3, H-4, and H-5), 2.14 (s, 3 H, H-8 methyl), 1.98 (d, $J = 7$ Hz, 3 H, H-6 methyl); IR (neat) 1745 and 1725 ($\text{C}=\text{O}$) cm^{-1} ; mass spectrum, m/e (relative intensity) 184 (M^+ , 4), 129 (45), 127 (23), 109 (26), 100 (35), 81 (89), 43 (100). The 2,4-DNP derivative of **2c** methyl ester melted at 122–123 °C.

Anal. Calcd for $\text{C}_{16}\text{H}_{20}\text{N}_4\text{O}_6$: C, 52.73; H, 5.53; N, 15.38. Found: C, 52.63; H, 5.47; N, 15.40.

1 α ,2 β ,5 α -Nepetonic Acid (2b) from 2c. To 30 mL of aqueous 10% NaOH under N_2 was added 320 mg (1.88 mmol) of **2c**. The solution was magnetically stirred for 2.5 h and then acidified with 10% HCl. The acidified mixture was extracted with ether and the ether extract was dried (Na_2SO_4) and evaporated under vacuum to give **2b** ($^1\text{H NMR}$ identical with that described above). Treatment of **2b** with CH_2N_2 and Kugelrohr distillation gave 300 mg (87% yield) of the methyl ester: $[\alpha]_D^{25} +35.2^\circ$ (c 3.4, CHCl_3); $^1\text{H NMR}$ and IR identical with those described above. The 2,4-DNP derivative of **2b** methyl ester was obtained in 60% yield: mp 78–80 °C; mixed with the 2,4-DNP methyl ester of **2b** obtained from **1b**, mp 76–78 °C; $[\alpha]_D +172^\circ$ (c 5.0, CHCl_3).

3 β ,3 $\alpha\beta$,6 $\alpha\beta$ -Nepetolactone (3c) and 3 α ,3 $\alpha\beta$,6 α ,6 $\alpha\beta$ -Nepetolactone (3d). A solution of 1.5 g (0.97 mmol) of **1c** in 75 mL of CH_3OH and 1.5 mL of pyridine was ozonized and freed of excess ozone as described for **1b**. The mixture was transferred to an ice

bath and 1.5-g portions of NaBH_4 , each in 15 mL of H_2O , were added to the stirred mixture, one immediately and others after 0.5, 1, 2, and 4 h. The mixture was then acidified with 10% HCl and extracted with ether. The ether extract was washed with NaHCO_3 solution, dried (MgSO_4), and concentrated to yield 500 mg (36% yield) of a mixture consisting of 65% **3c** and 35% **3d**, as indicated by GC analysis. Lactones **3c** and **3d** were separated by preparative GC. Lactone **3c**: bp 60–65 °C (0.05 mm); $[\alpha]_D^{24} +15.1^\circ$ (c 2.4, CHCl_3); $^1\text{H NMR}$ (CDCl_3) δ 4.15 (m, 1 H, H-3), 3.08–1.40 (7 H, CH and CH_2), 1.37 (d, $J = 6$ Hz, 3 H, H-7), 1.17 (d, $J = 7$ Hz, 3 H, H-8); mass spectrum, m/e (relative intensity) 154 (M^+ , 23), 99 (70), 94 (40), 82 (76), 81 (93), 67 (100).

Anal. Calcd for $\text{C}_9\text{H}_{14}\text{O}_2$: C, 70.10; H, 9.15. Found: C, 69.90; H, 9.26.

Lactone **3d**: bp 60–65 °C (0.05 mm); $[\alpha]_D^{24} +78.9^\circ$ (c 1.0, CHCl_3); $^1\text{H NMR}$ (CDCl_3) δ 4.69 (m, 1 H, H-3), 3.12–1.40 (7 H, CH and CH_2), 1.37 (d, $J = 6$ Hz, 3 H, H-7), 1.24 (d, $J = 7$ Hz, 3 H, H-8); mass spectrum, m/e (relative intensity) 154 (M^+ , 16), 99 (48), 95 (45), 82 (75), 81 (100), 67 (99).

Anal. Calcd for $\text{C}_9\text{H}_{14}\text{O}_2$: C, 70.10; H, 9.15. Found: C, 70.25; H, 9.15.

3 α ,3 $\alpha\alpha$,6 α ,6 $\alpha\alpha$ -Nepetolactone (3a) and 3 β ,3 $\alpha\alpha$,6 α ,6 $\alpha\alpha$ -Nepetolactone (3b). Lactones **3a** and **3b** were prepared from **1a** by the above procedure in 55% yield. Gas chromatographic analysis indicated the mixture to consist of 42% **3a** and 58% **3b**. Preparative GC gave the pure individual lactones. Lactone **3a**: $^1\text{H NMR}$ (CDCl_3) δ 4.31 (m, 1 H, H-3), 2.70–1.30 (7 H, CH and CH_2), 1.36 (d, $J = 6$ Hz, 3 H, H-7), 1.11 (d, $J = 6$ Hz, 3 H, H-8); mass spectrum, m/e (relative intensity) 154 (M^+ , 2), 139 (17), 95 (44), 82 (50), 81 (100), 67 (69). Lactone **3b**: $^1\text{H NMR}$ (CDCl_3) δ 4.60 (m, 1 H, H-3), 2.90–1.30 (7 H, CH and CH_2), 1.30 (d, $J = 6$ Hz, 3 H, H-7), 1.07 (d, $J = 6$ Hz, 3 H, H-9); mass spectrum, m/e (relative intensity) 154 (M^+ , 1), 99 (70), 95 (30), 94 (40), 82 (40), 81 (100), 67 (53).

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Registry No. **1a**, 21651-62-7; **1b**, 17257-15-7; **1c**, 21651-53-6; **2b**, 21651-52-5; **2b** methyl ester, 6890-02-4; **2b** methyl ester 2,4-DNP derivative, 74366-00-0; **2c**, 58801-35-7; **2c** methyl ester, 74410-38-1; **2c** methyl ester 2,4-DNP derivative, 74410-39-2; **3a**, 74410-40-5; **3b**, 74410-41-6; **3c**, 58845-57-1; **3d**, 58845-58-2.

New Diterpenoids from the Soft Corals *Xenia macrospiculata* and *Xenia obscuronata*

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Sixteen diterpenoids have been isolated from two soft corals of the Red Sea, *Xenia macrospiculata* and *X. obscuronata*. These xenia diterpenoids, many of them novel and reported for the first time, are divided into three subgroups: the xenicins [xeniculin (2), 9-deacetoxy-14,15-deepoxyxeniculin (3), and 9-deacetoxy-14,15-deepoxyxeniculin 7,8-epoxide (4)], the xeniolides [xeniolide-A (6), xeniolide-B (7a), xeniolide-B 9-acetate (7b), 7,8-epoxyxeniolide-B (8), and xenialactol (9a)], and the xeniaphyllanes [xeniaphyllenol (11), 4,5-epoxyxeniaphyllenol (12), isoxeniaphyllenol (13), 4,5-epoxyisoxeniaphyllenol (14), 14,15-xeniaphyllandiol (15a), xeniaphyllandiol 14-acetate (15b), and 4,5-epoxy-14,15-xeniaphyllandiol (16)]. The structures were elucidated from spectral data and chemical transformations. The ^{13}C NMR spectra are fully assigned, and their application to the determination of the xenia diterpenoids is discussed. In addition to the above compounds, another diterpenoid, obscuronatin (23), was isolated and its structure elucidated.

The soft corals are known to be a source of many different diterpenoids.¹ Many of these diterpenoids belong

to a single chemical class, the cembranoids.^{2,3} This report deals with a new group of compounds, the xenia di-